

Datasheet

SERAplex NTA

Defined Serum Substitute for Adherent Cells

Product	Description	Catalogue-No.	Size
SERAplex NTA	Defined serum substitute for adherent cells	ST04-95070 ST04-95700 ST04-95750	50 ml 100 ml 500 ml

Product description

SERAplex NTA is a ready-to-use, chemically defined serum substitute for the cultivation of adherent cells under serum-free conditions or to significantly reduce the amount of serum in cell culture. It supports the growth of many adherent cell types in an optimum manner without any extra handling compared to serum.

Storage conditions

Storage: -20°C (in the dark)

Stability: 2 years from date of production

Size: 50 ml, 100 ml, 500 ml, other sizes on request

Composition

SERAplex NTA contains purified proteins, lipids, salts, amino acids, trace elements, attachment factors and hormones in an optimized formulation. SERAplex NTA contains no growth factors, undefined hydrolysates or peptones.

Suitability

SERAplex NTA is suitable for the cultivation of a variety of adherent cells under serum-free culture conditions (please see figure 1) or to reduce the necessary FBS amount in cell culture.

Effect of SERAplex NTA in different cell lines

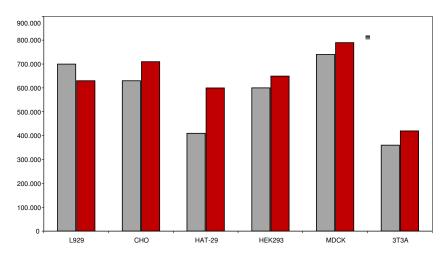


Fig. 1: Efficiency and growth stimulation of SERAplex NTA compared to FBS (10% each in DMEM/F12) at 7 day culture in different cell types

FBS

SERAplex NTA



Special advantages

SERAplex NTA is designed to replace or to reduce serum in the cell culture in a very simple manner. In most cases there is no need to change the basal medium. As SERAplex NTA is fully chemically defined and contains no peptones or hydrolysates, lot testing is no more necessary. It also allows high reproducibility and a simplified downstream process. SERAplex NTA contains no growth factors and enables defined proliferation and differentiation of stem cells. Characterization studies of growth factors will obtain more reproducible and clearer results. SERAplex NTA is also useful to develop sensitive cell-based *in vitro* tests and coculture procedures. For cell lines which require specific growth factors these should be added in a concentration as previously used.

Instructions for use

SERAplex NTA can be stored and used in the same manner as serum.

- Thaw SERAplex at maximum 37 °C. Please avoid repeated freeze-thaw cycles!
- <u>To replace serum</u>: Use the same basal medium and the same concentration of SERAplex as FBS. The performance can be further improved by optimizing the concentration of SERAplex or modifying/changing the basal medium^a (Table 1)
- <u>To reduce serum concentration</u>: Use the same basal medium and add the same amount of SERAplex as the reduced amount of serum, until the minimal necessary concentration of FBS is found (1 to 2.5 % in most cases). The performance can be further improved by optimizing the the concentration of SERAplex or modifying/changing the basal medium^a (also see adaptation instruction and table 1).
- Recommended inoculation cell density: 5.000 20.000 cells/cm².
- Solve cells as usual from the cell culture vessel (e.g. 0.25% trypsin inhibitor, Cat.No. ST10-033100 or Accutase[®], Cat.No. ST10-21100). Once the cells have become round and detach from the surface inactivate trypsin with trypsin inhibitor (Cat.No. ST10-034100): Simply resuspend cells in about 1 ml trypsin inhibitor solution for every ml of trypsin solution used for dissociation. Note that Accutase[®] does not need to be inhibited.

Depending on the cell type, some differences in morphology or proliferation rate may be observed with varying standard media. Most applications were performed with DMEM and DMEM/F12 for adherent cells. Make sure that L-glutamine is present in sufficient quantity. The optimal SERAplex NTA concentration should be determined for each cell line. Tests can be started at a SERAplex NTA concentration of 10%, as with most cells the best results were obtained at this concentration.

Please note: For more demanding cells an adaptation to SERAplex NTA may be necessary.



Adaptation instructions for SERAplex NTA

Precondition for a successful transition are vital cells (trypan blue exclusion staining), which should be harvested in the logarithmic growth phase. If 10% FBS was used in the original protocol,

Step 1: 7.5 % FBS + 2.5 % SERAplex

- Seed cells at 5 x 10³ 20 x 10³ cells/cm².
- Observe cells under a microscope, at about 90 % confluence passage the cells for another 2-3 passages.

If normal growth is obtained transfer cells into:

Step 2:5 % FBS + 5 % SERAplex

- Seed cells at 5 x 10³ 20 x 10³ cells/cm².
- Observe cells under a microscope, at about 90 % confluence passage the cells for another 2-3 passages.

If normal growth is obtained transfer cells into:

Step 3: 2.5 % FBS + 7.5 % SERAplex

- Seed cells at 5 x 10³ 20 x 10³ cells/cm².
- Observe cells under a microscope, at about 90 % confluence passage the cells for another 2-3 passages.

If normal growth is obtained transfer cells into:

Step 4: 1 % FBS + 9 % SERAplex

- Seed cells at 5x10⁴ 10x10⁵ cells/ml.
- Observe cells under a microscope, at about 90 % confluence passage the cells for another 2-3 passages.

If normal growth is obtained transfer cells into:

Step 5: 10 % SERAplex

• Seed cells at 5 x 10³ – 20 x 10³ cells/cm².

Observe cells under a microscope.

For some cells an adaptation to serum-free conditions is difficult to reach or even impossible.

The following measures may help to facilitate a successful adaptation:

- Reseeding with a higher cell amount (about 2x to 4x of the usual cell density).
- Addition of growth factors (if known, which factors have a positive effect on the relevant cells).
- Coating the culture dishes or flasks with attachment factors (e.g. fibronectin, laminin, collagen, gelatine, etc).
- Change the basal medium. Note: A change of the basal medium to a richer or more complex formulation may be all that is needed to achieve growth in serum free condition.



Table 1: Comparison of Cell Growth in 10% SERAplex NTA in different Basal Media versus Cell Growth in 10% FBS in different Basal Media

Cell Line	Origin	Basal Medium	Percentage of Growth 10% SERAplex NTA	Percentage of Growth 10% FBS	
HEK 293 T	Renal cells, human	DMEM/F12	105%	100%	
	embryonic	alpha-MEM	76%	100%	
		DMEM	62%		
MDCK	Renal cells, canine	DMEM/F12	102%	100%	
		McCoy's 5A	91%	100%	
		alpha-MEM	106%		
MDBK	Renal cells, bovine	RPMI 1640	122%	100%	
		McCoy's 5A	135%	100%	
		DMEM	131%		
L 929	Fibroblasts, mouse	DMEM	97%	100%	
		RPMI 1640	78%	100%	
		Ham's F-12	128%		
HT-29	Colon Carcinoma,	IMDM	108%	1000/	
	human	DMEM/F12	98%	100%	
		alpha-MEM	96%		
HeLa S3	Cervix carcinoma	Glasgow MEM	106%	100%	
	epithel, human	IMDM	72%		
	•	EMEM	100%		
СНО	Ovarial cells epithel,	DMEM/F12	106%	1000/	
	Chinese hamster	IMDM	97%	100%	
		alpha-MEM	82%		
CHO-Luc	Ovarial cells epithel,	IMDM	86%	100%	
	Chinese hamster,	DMEM	97%		
	transfected	alpha-MEM	84%		
3T3A	Fibroblasts, mouse	RPMI 1640	98%	1000/	
		McCoy's 5A	72%	100%	
		DMEM/F12	97%		
MCF-7	Mammary	Ham's F-12	292%	1000/	
	carcinoma, human	DMEM/F12	176%	100%	
		RPMI 1640	214%		
RAW 264.7	Macrophages,	McCoy's 5A	40%	1000/	
	mouse	DMEM/F12	67%	100%	
		alpha-MEM	38%		

Technical support

For technical support, questions or remarks please contact your local PAN-Seratech partner or the technical department of PAN-Seratech via email (info@pan-seratech.com) or phone +49-8543-601630.

FOR RESEARCH USE ONLY! Not approved for human or animal diagnostic or therapeutic procedures.

^a As a basal medium, standard media such as RPMI 1640, DMEM (high or low glucose), DMEM/F12, IMDM etc. can be used. Make sure that L-glutamine is present in sufficient quantity (supplement L-glutamine as needed).